

Histology Sampling Protocol - AquaTechnics

PURPOSE: Capture samples before, during and after times of oyster mortality and/or stress for professional, histological diagnostics

General Guidelines:

1. Collect 3-5 samples as oyster progress
2. Send samples after mortality event. Not all samples will need to be shipped/processed (e.g. If a mortality event does not occur)
3. Send in up to 20 samples per year

Supplies: 30 - 50 ml Nalgene containers, Seawater Formalin (10% Formalin solution (10% Full strength formalin [37%] + 90% filtered seawater [use at least 1 um filtered seawater, or smaller nominal pore filter equivalent])).

Sample Prep:

1. Add seed to sample tube as specified in the table below for specific seed size.
2. Add fixative (10x seed volume) to sample tube. For example, if you put 3 mL of packed 4 mm seed in the tube, bring the volume up at least 30 mL with fixative.
3. Screw cap on tightly. Samples can be stored in this manner for several weeks.
Note: samples must be fixed at least 2 days prior to shipping

SHELL LENGTH	SAMPLE METHOD	#/VOLUME OF SEED
1 mm – 4 mm	Pack seed into sample tube	3 or 4 mL
4 mm – 7 mm	Count into sample tube	At least 30
> 6-8 mm	Shuck oysters, add meat only	30

Ready to Ship:

1. Drain off most of the fixative when you are ready to ship, but leave samples moist.
2. Make sure the screw caps are on tight and the tubes are clearly labelled
3. Double zip lock the container
4. Make sure you include a paper log describing the samples or email the log. Make sure the log numbers clearly correspond to the sample vial numbers.
5. Please send a heads up email that you are shipping samples to:
ralph@aquatechnics.com
6. Ship samples by UPS Ground to:

AquaTechnics
455 West Bell Street
Sequim, WA 98382
(Tel: 360-681-3122)

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Contact: For more information about this project, protocols, or how to request supplies contact Pacific Shellfish Institute (3/2018).



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